

# Protocol: Flow Cytometric Staining with Cell Surface Abs

## Steps:

1. Sample collection and dispensing:
  - a. Whole blood: dispense 50-200  $\mu\text{L}$  per sample.
  - b. Fresh PBMC: resuspend at  $5-10 \times 10^6$  viable lymphocytes per mL in wash buffer or cRPMI, dispense 50-200  $\mu\text{L}$  per sample.
  - c. Cryopreserved PBMC: Follow PBMC Cryopreservation protocol, resuspend at  $5-10 \times 10^6$  viable lymphocytes per mL in warm cRPMI, dispense 50-200  $\mu\text{L}$  per sample.
2. Staining:
  1. Add appropriate titers of all antibodies, incubate for 30-60 m at room temperature in the dark. If possible, vortex or otherwise agitate occasionally during staining.
3. Washing:
  1. Whole blood: add appropriate volume of an erythrocyte lysing buffer (e.g., BD PharmLyse or BD FACS Lysing Solution). Follow manufacturer's directions for incubation and washing.
  2. PBMC: add wash buffer to at least 200  $\mu\text{L}$ , centrifuge at  $250 \times G$  for 5 m at room temperature. Repeat wash a second time for optimal results.
4. Final fixation (optional):
  - a. For assays with Cy7 tandems, but NOT AmCyan: Resuspend in at least 200  $\mu\text{L}$  of BD Stabilizing Fixative and store at 4C in the dark. Acquire within 24 h if possible.
  - b. For other assays: Resuspend in at least 200  $\mu\text{L}$  of 0.5-2% paraformaldehyde in PBS and store at 4C in the dark. Acquire within 24 h if possible.

## Notes:

1. Staining can be done in 5-mL FACS tubes, or various types of 96-well conical or round-bottom plates.
2. Antibodies should be titrated for the intended application (sample type, surface vs intracellular staining, number of cells, staining vessel/volume, time, and temperature). Choose the titer point that maximizes signal:noise.
3. It can be convenient to create a master mix of all antibodies in wash buffer so that, for example, 50  $\mu\text{L}$  are added to each 50  $\mu\text{L}$  cell sample (for a total staining volume of 100  $\mu\text{L}$ ).
4. Most antibodies work well when stained for 30 m at room temperature. Longer times (up to 60 m) can improve weak staining, as can agitation during Ab incubation. Occasional Abs may work better when stained on ice.
5. After fixation, cells become more bouyant, and need to be centrifuged at higher G force ( $500 \times G$ ).
6. Fixation of stained samples can degrade staining of some sensitive markers. Resuspending in wash buffer is preferable, even if cells will be stored overnight prior to acquisition (keep dark at 4C).

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## Stock Solution Recommendations:

| <b>Solution</b> | <b>Stock Concentration</b>            | <b>Intermediate Dilution</b> | <b>Final Concentration</b> |
|-----------------|---------------------------------------|------------------------------|----------------------------|
| Wash Buffer     | 0.5% BSA+0.1% NaN <sub>3</sub> in PBS | None                         | Use neat                   |
| Final fixative  | 0.5% paraformaldehyde in PBS          | None                         | Use neat                   |